



Why does epithelia display heterogeneity? Bridging physical and biological concepts

Medhavi Vishwakarma^{1,2} • Jacopo Di Russo^{2,3}

Received: 18 July 2019 / Accepted: 27 August 2019 / Published online: 7 September 2019

© International Union for Pure and Applied Biophysics (IUPAB) and Springer-Verlag GmbH Germany, part of Springer Nature 2019

Epithelial cells construct inner and outer linings of our organs and function as physical barriers, thus, protecting the underlying tissue from infections, dehydration, and also aiding in efficient absorption of nutrients and gases (Alberts 2008). Cells within the epithelia perform these tasks, being jammed at their place while also making sure that epithelial homeostasis is maintained, failing in which can be potentially fatal for the tissue (Macara et al. 2014). Interestingly, the same cells can unjam and flow almost like a fluid during physiological and pathological situations such as organ development, wound healing and cancer metastasis (Friedl and Gilmour 2009; Mongera et al. 2018; Park et al. 2016; Sadati et al. 2013; Scarpa and Mayor 2016). In such situations, cells, rather than moving individually, migrate as a group in various patterns (Haeger et al. 2015; Petitjean et al. 2010; Poujade et al. 2007; Rorth 2012; Tarle et al. 2015). Reductionist view holds that such cooperative cellular events are mediated at the level of cell-cell interactions where local signals are translated into physical forces (such as those generated in the cellular cytoskeleton and those exerted across cell-cell junctions), which are then translated into cell motility (Das et al. 2015; Keller 2012; Ladoux and Mège 2017; Trepat et al. 2009). Such physical forces are believed to be fundamental to biological form and function but have remained hidden until recently when experimental methods are finally making them visible (Angelini et al. 2010; Angelini et al. 2011; Edwards and

Schwarz 2011; Malinverno et al. 2017; Sabass et al. 2008; Schwarz and Soine 2015; Sunyer et al. 2016; Tambe et al. 2011; Trepat and Fredberg 2011). Furthermore, recent advances in mathematical biology have also led to the development of models that can predict various parameters of epithelial behaviour in both jammed and unjammed states (Edwards and Schwarz 2011; Garcia et al. 2015; Henkes et al. 2011; Mark et al. 2010; Mehes and Vicsek 2014; Sepulveda et al. 2013; Steinberg 2007). Together, these studies have revealed unpredicted behaviour of epithelial tissues and are beginning to explain why cells jam and unjam, and how collective cell behaviour is orchestrated. Since many excellent reviews have been written on the topic (Friedl and Gilmour 2009; Haeger et al. 2015; Merkel and Manning 2017; Park et al. 2016; Park and Fredberg 2016; Pegoraro et al. 2016; Sadati et al. 2013), here, we will only briefly describe the heterogeneous nature of the jamming transition from the physical perspective and focus mainly on its implications in regulating epithelial functionality while also taking into account the inherent biological heterogeneity present within the epithelium.

Jamming transition and dynamic heterogeneity

Ongoing cell divisions, apoptosis and cell mingling make the epithelia a highly dynamic place (Al-Hussaini et al. 2016; Christ et al. 1990; Gardner 1986; Macara et al. 2014). Interestingly, monolayer stress profiles of such epithelial layers reveal dynamic heterogeneity, with intercellular stress displaying stochasticity in space and time, meaning that stress is tied neither to any particular position nor to any particular cell within the monolayer (Angelini et al. 2010, 2011; Garrahan 2011; Tambe et al. 2011). Topography of these intercellular forces, at any given instant, can be compared with a rugged landscape, similar to that of a mountain range, where peaks arise from cooperation between tens of cells pulling together (Tambe et al. 2011) (Fig. 1a). Interestingly, cell

✉ Medhavi Vishwakarma
m.vishwakarma@bristol.ac.uk

✉ Jacopo Di Russo
jdirusso@ukaachen.de

¹ School of Cellular and Molecular Medicine, University Walk, University of Bristol, Bristol BS1 8TD, UK

² Max Planck Institute for Medical Research, Jahnstrasse 29, 69120 Heidelberg, Germany

³ Interdisciplinary Centre for Clinical Research, RWTH Aachen University, Pauwelstrasse 30, 52074 Aachen, Germany

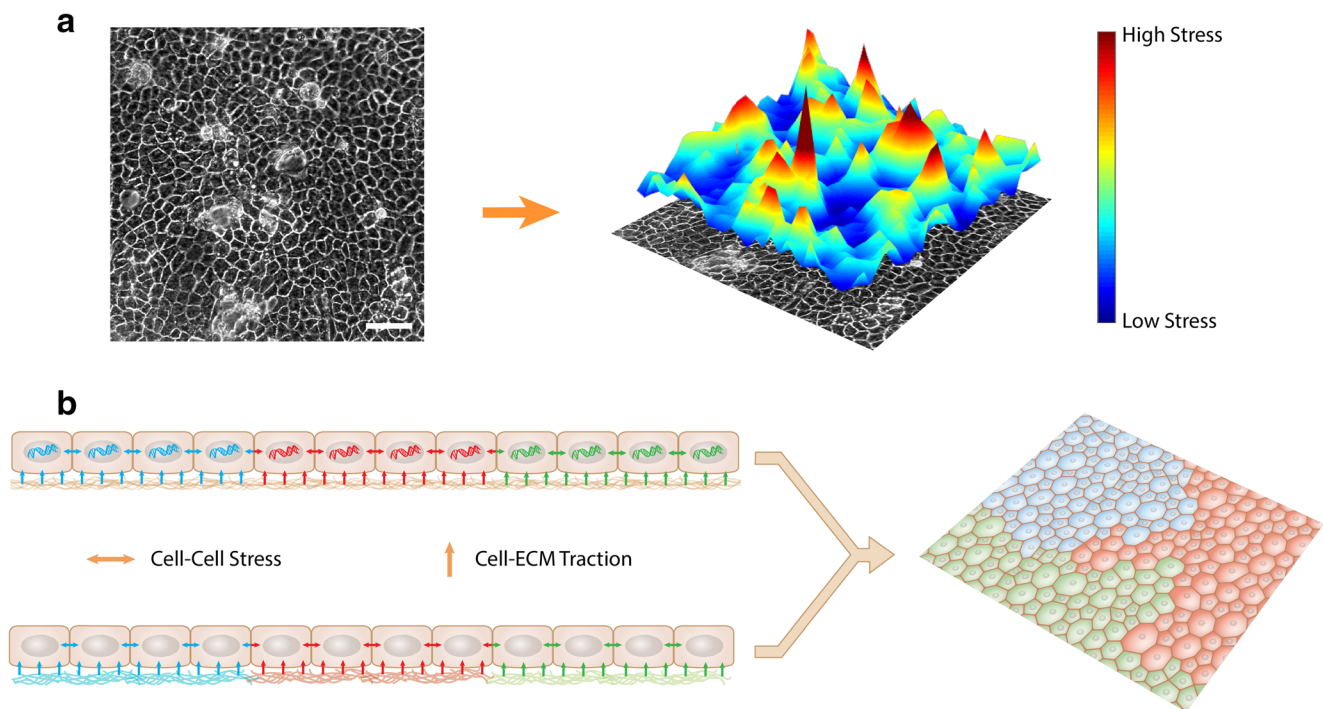


Fig. 1 **a** The intercellular stress profile in a confluent epithelial monolayer of canine kidney epithelial cells (MDCK) reveal a rugged stress profile at a given time point. Scale bar is 50 μm . **b** Cellular heterogeneities can arise from genetic differences or differential regulation of protein expression which are also influenced by external cues such as

ECM components. In addition, heterogeneous clones in epithelia might differ in their mechanical properties, having different levels of adhesion forces (cell-cell stresses and cell-ECM tractions), thus impacting on physical nature of epithelia

density also plays a key role in regulating dynamic heterogeneity; i.e. when cells start to crowd, their movement becomes arrested and zones of cooperativity grow bigger (Angelini et al. 2011). Such a scenario is intriguingly analogous to glass transition within a supercooled fluid or dense particulate matter in which a non-equilibrium jammed state is reached by cooling, crowding or by decreasing applied load (Debenedetti and Stillinger 2001; Mattsson et al. 2009; Mayer et al. 2008; Nagel 1998; Trappe et al. 2001). Hallmarks similar to glass transition (spontaneous intermittent fluctuations, dynamic heterogeneity, cooperativity and kinetic arrest) are observed by epithelial cell monolayer, wherein the dynamical arrest is caused upon crowding and depends upon parameters such as active motility, cellular forces, cell shape and applied stress. When these parameters are comprehended in a jamming phase diagram (Nagel 1998; Sadati et al. 2013; Trappe et al. 2001), predictions on epithelial physical behaviour can be made. For instance, as intercellular adhesion or crowding progressively increases, cell motility and rearrangement would become rare and therefore, cooperativity would increase, leading to a topologically frozen epithelium (Sadati et al. 2013). Subsequently, then, the question is what the extension of jamming at homeostasis should be that allows the epithelia to achieve their vital physiological functions such as regulating homeostasis and orchestrating collective cell migration.

Physiological relevance of heterogeneity

The ability of epithelial cells to dynamically remodel their surroundings as well as their own cytoskeleton in response to external cues such as damage or mechanical stresses is known to provide a mechanical resilience to epithelial tissues (Khalilgharibi et al. 2019; Trepap and Sahai 2018). Recent studies are suggestive of the hypothesis that, by maintaining a striking balance between jammed and unjammed phases, the epithelial monolayer might have evolved to attain such resilience, by virtue of which, it can efficiently undergo switch-like changes required for physiological functions (Park et al. 2015; Sadati et al. 2014; Saw et al. 2017; Vishwakarma et al. 2018). For instance, a recent study demonstrates that cooperative forces owing to dynamic heterogeneity control the selection as well as frequency of leader cells which guide collective migration during wound healing (Vishwakarma et al. 2018). Another study demonstrates that hot spots of compressive stresses within the epithelial monolayer induce topological defects that subsequently lead to local cell extrusion (Saw et al. 2017). Since hot spots of compressive stresses build up regions of multicellular cooperation (Tambe et al. 2011) which show density dependence (Angelini et al. 2011), efficient cell extrusion for regulating tissue homeostasis would intuitively require the right extent of cell packing. Such extrusion events are important, due to their relevance not only in regulating cell

density during epithelial homeostasis (Fadul and Rosenblatt 2018; Gudipaty et al. 2018) but also in removing aberrant or tumour cells via a mechanism described as cell competition, by virtue of which, epithelia gain the ability to defend itself against cancer (Kajita and Fujita 2015; Wagstaff et al. 2013). Understanding physiological relevance and extent of jamming in epithelia becomes even more important in tissues that are naturally subjected to elevated levels of stress, such as lung epithelium which goes through cyclic breathing stress and, therefore, tissue plasticity plays an important role in maintaining its integrity, especially during lung injury (Frank and Matthay 2003).

The physical heterogeneity described above is most likely to be influenced by the existing innate biological heterogeneities in the epithelia which are associated with variations in genome or protein expression patterns (Fig. 1b). A known outcome of this genetic variability is the somatic mosaicism that leads to the presence of multiple cell clones within an adult tissue. Somatic mosaicism can originate from epigenetics events (Rakyan et al. 2002; Sutherland et al. 2000) such as, for instance, the inactivation of one of the X-chromosomes in females (Rakyan et al. 2002) or from mobile DNA elements such as retrotransposons (Beck et al. 2011; De 2011). A classic example of somatic mosaicism can be observed in the skin with the presence of mosaicisms in the pigmentation known as *café-au-lait* spots (De 2011; Rawles 1947). In addition to these genetic differences, differential regulation of proteins expression induced by external cues, such as extracellular matrix (ECM), can also create cellular heterogeneity within the epithelial layer (Fig. 1b). The importance of such heterogeneity in regulating tissue homeostasis has been shown in the basal layer of esophageal epithelium containing stem cells responsible for tissue renewal (DeWard et al. 2014). It has been shown that, in this layer, population of stem cells has heterogenous proliferation rates which are distinguishable by the expression of specific cell-surface markers such as the laminin receptor integrin $\alpha 6\beta 4$. Here, the involvement of laminins, major components of extracellular matrix, suggests the importance of cell-ECM adhesion in maintaining cellular heterogeneity and, subsequently, in regulating tissue homeostasis (DeWard et al. 2014). Interestingly, cellular heterogeneity dictated by differential laminin expression has also been shown to be involved in regulating functionality of endothelial cells. For example, the extracellular matrix of endothelium in postcapillary venules consists of areas of high and low expression of the laminin 511 isoform compared with the capillaries where the expression of laminin 511 is homogeneous (Di Russo et al. 2017; Sixt et al. 2001). Such differential distribution of laminin controls endothelial cell junction tightness, thereby dictating the location of leucocytes extravasation through the blood-brain barrier which occurs only in low laminin 511 regions (Sixt et al. 2001; Song et al. 2017). In addition to the biochemical composition of ECM, its topography

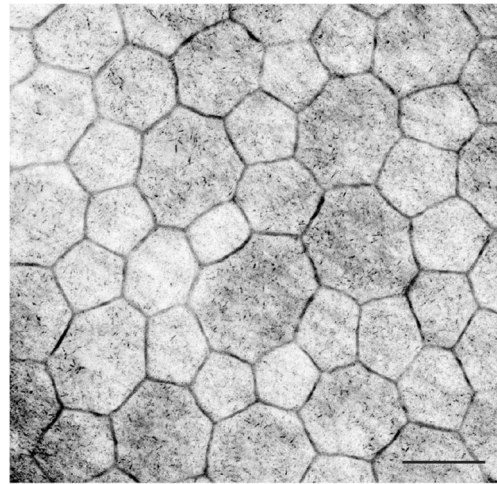


Fig. 2 Immunofluorescent staining of *en face* preparation of murine retinal pigment epithelium for filamentous actin reveals highly heterogeneous character of this epithelium. To be noted is the postmitotic nature of these epithelial cells that exclude correlation of cell size with the cell cycle. Scale bar is 20 μ m

has also been shown to control the heterogeneity of epithelial cells. Recently, an elegant experimental setting using undulated elastomer surfaces revealed the effect of ECM topography on heterogeneity of keratinocytes (Mobasserri et al. 2019). After seeding primary keratinocyte on the surfaces, the monolayer assembled within a range of cellular stiffness, cell-cell adhesion forces and acto-myosin contractility levels. The results provided new insights into the possible heterogeneous control of keratinocytes proliferation rates by the topography of the dermal ECM during ageing and inflammation (Mobasserri et al. 2019). Differential ECM expression also impacts on the aetiology of retinal degenerative disease, i.e. age-related macular degeneration. The early stage of the disease is characterized by high level of ECM accumulation known as *drusen* that occurs between the retinal pigment epithelium and the underlying Bruch's membrane (Coleman et al. 2008). Drusen formation is a common age effect, but only the accumulation of high number of large drusen ($> 63 \mu$ m in diameter) correlates with epithelium degeneration and photoreceptor detachment (Coleman et al. 2008). Since the retina pigment epithelium presents a very high heterogeneity in cell shape (Fig. 2), protein synthesis and granule accumulation, it is tempting to speculate that this diversity of cell shape might also correspond to high heterogeneity in monolayer tensions and, therefore, might control drusen formation and their growth.

Conclusion

Even though physical and biological heterogeneities are currently known to be distinct, they are likely to be interactive and interdependent. Local cellular heterogeneity might

influence the mechanical properties of epithelia, its ability to transduce forces and, hence, the nature of physical heterogeneity. Recent technological advancements in biophysics, cell biology and mathematical biology have now made it possible to analyse the physics and biology of the epithelia within the same framework. Such approaches allow us to attain a more comprehensive understanding on epithelial physiology and would subsequently require devising new treatment strategies for epithelial degenerative diseases.

Acknowledgements We appreciate the support of the interdisciplinary centre for clinical research (IZKF) and the institute for molecular and cellular anatomy (MOCA) at RWTH Aachen, as well as the school of cellular and molecular medicine at the University of Bristol. M.V. and J.D.R. are guest scientists at the Max Planck Institute for Medical Research in Heidelberg, Germany. We thank Adam Breitscheidel for his support in the graphic design and Natalia Simon for proofreading.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- Alberts B (2008) Molecular biology of the cell. Reference edition, 5th edn. Garland Science, New York
- Al-Hussaini H, Kilarkaje N, Shahabi G, Al-Mulla F (2016) Proliferation and migration of peripheral retinal pigment epithelial cells are associated with the upregulation of wingless-related integration and bone morphogenetic protein signaling in Dark Agouti rats. *Med Princ Pract* 25:408–416. <https://doi.org/10.1159/000446480>
- Angelini TE, Hannezo E, Trepas X, Fredberg JJ, Weitz DA (2010) Cell migration driven by cooperative substrate deformation patterns. *Phys Rev Lett* 104:168104. <https://doi.org/10.1103/PhysRevLett.104.168104>
- Angelini TE, Hannezo E, Trepas X, Marquez M, Fredberg JJ, Weitz DA (2011) Glass-like dynamics of collective cell migration. *Proc Natl Acad Sci U S A* 108:4714–4719. <https://doi.org/10.1073/pnas.1010059108>
- Beck CR, Garcia-Perez J, Badge RM, Moran JV (2011) LINE-1 elements in structural variation and disease. *Annu Rev Genomics Hum Genet* 12:187–215. <https://doi.org/10.1146/annurev-genom-082509-141802>
- Christ B, Poelmann RE, Mentink MMT, Groot G-dAC (1990) Vascular endothelial cells migrate centripetally within embryonic arteries. *Anat Embryol* 181:333–339. <https://doi.org/10.1007/BF00186905>
- Coleman HR, Chan C-C, Ferris FL, Chew EY (2008) Age-related macular degeneration. *Lancet* 372:1835–1845. [https://doi.org/10.1016/S0140-6736\(08\)61759-6](https://doi.org/10.1016/S0140-6736(08)61759-6)
- Das T, Safferling K, Rausch S, Grabe N, Boehm H, Spatz JP (2015) A molecular mechanotransduction pathway regulates collective migration of epithelial cells. *Nat Cell Biol* 17:276–287. <https://doi.org/10.1038/ncb3115>
- De S (2011) Somatic mosaicism in healthy human tissues. *Trends Genet* 27:217–223. <https://doi.org/10.1016/j.tig.2011.03.002>
- Debenedetti PG, Stillinger FH (2001) Supercooled liquids and the glass transition. *Nature* 410:259–267. <https://doi.org/10.1038/35065704>
- DeWard AD, Cramer J, Lagasse E (2014) Cellular heterogeneity in the mouse esophagus implicates the presence of a nonquiescent epithelial stem cell Population. *Cell Rep* 9:701–711. <https://doi.org/10.1016/j.celrep.2014.09.027>
- Di Russo J et al (2017) Vascular laminins in physiology and pathology. *Matrix Biol* 57–58:140–148. <https://doi.org/10.1016/j.matbio.2016.06.008>
- Edwards CM, Schwarz US (2011) Force localization in contracting cell layers. *Phys Rev Lett* 107:128101. <https://doi.org/10.1103/PhysRevLett.107.128101>
- Fadul J, Rosenblatt J (2018) The forces and fates of extruding cells. *Curr Opin Cell Biol* 54:66–71. <https://doi.org/10.1016/jceb.2018.04.007>
- Frank JA, Matthay MA (2003) Science review: mechanisms of ventilator-induced injury. *Crit Care (London, England)* 7:233–241. <https://doi.org/10.1186/cc1829>
- Friedl P, Gilmour D (2009) Collective cell migration in morphogenesis, regeneration and cancer. *Nat Rev Mol Cell Biol* 10:445–457. <https://doi.org/10.1038/nrm2720>
- Garcia S, Hannezo E, Elgeti J, Joanny JF, Silberzan P, Gov NS (2015) Physics of active jamming during collective cellular motion in a monolayer. *Proc Natl Acad Sci U S A* 112:15314–15319. <https://doi.org/10.1073/pnas.1510973112>
- Gardner RL (1986) Cell mingling during mammalian embryogenesis. *J Cell Sci Suppl* 4:337–356
- Garrahan JP (2011) Dynamic heterogeneity comes to life. *Proc Natl Acad Sci* 108:4701–4702. <https://doi.org/10.1073/pnas.1101436108>
- Gudipaty SA, Conner CM, Rosenblatt J, Montell DJ (2018) Unconventional ways to live and die: cell death and survival in development, homeostasis, and disease. *Annu Rev Cell Dev Biol* 34:311–332. <https://doi.org/10.1146/annurev-cellbio-100616-060748>
- Haeger A, Wolf K, Zegers MM, Friedl P (2015) Collective cell migration: guidance principles and hierarchies. *Trends Cell Biol* 25:556–566. <https://doi.org/10.1016/j.tcb.2015.06.003>
- Henkes S, Fily Y, Marchetti MC (2011) Active jamming: self-propelled soft particles at high density. *Phys Rev E Stat Nonlinear Soft Matter Phys* 84:040301. <https://doi.org/10.1103/PhysRevE.84.040301>
- Kajita M, Fujita Y (2015) EDAC: epithelial defence against cancer-cell competition between normal and transformed epithelial cells in mammals. *J Biochem* 158:15–23. <https://doi.org/10.1093/jb/mvv050>
- Keller R (2012) Developmental biology) Physical biology returns to morphogenesis. *Science* 338:201–203. <https://doi.org/10.1126/science.1230718>
- Khalilgharibi N et al. (2019) Stress relaxation in epithelial monolayers is controlled by the actomyosin cortex. *Nat Phys* 1–9. <https://doi.org/10.1038/s41567-019-0516-6>
- Ladoux B, Mège R-MM (2017) Mechanobiology of collective cell behaviours. *Nat Rev Mol Cell Biol* 18:743–757. <https://doi.org/10.1038/nrm.2017.98>
- Macara IG, Guyer R, Richardson G, Huo Y, Ahmed SM (2014) Epithelial homeostasis. *Curr Biol* 24:R815–R825. <https://doi.org/10.1016/j.cub.2014.06.068>
- Malinverno C et al (2017) Endocytic reawakening of motility in jammed epithelia. *Nat Mater* 16:587–596. <https://doi.org/10.1038/nmat4848>
- Mark S, Shlomovitz R, Gov NS, Poujade M, Grasland-Mongrain E, Silberzan P (2010) Physical model of the dynamic instability in an expanding cell culture. *Biophys J* 98:361–370. <https://doi.org/10.1016/j.bpj.2009.10.022>
- Mattsson J, Wyss HM, Fernandez-Nieves A, Miyazaki K, Hu Z, Reichman DR, Weitz DA (2009) Soft colloids make strong glasses. *Nature* 462:83–86. <https://doi.org/10.1038/nature08457>
- Mayer C et al (2008) Asymmetric caging in soft colloidal mixtures. *Nat Mater* 7:780–784. <https://doi.org/10.1038/nmat2286>
- Mehes E, Vicsek T (2014) Collective motion of cells: from experiments to models. *Integr Biol (Camb)* 6:831–854. <https://doi.org/10.1039/c4ib00115j>

- Merkel M, Manning ML (2017) Using cell deformation and motion to predict forces and collective behavior in morphogenesis. *Semin Cell Dev Biol* 67:161–169. <https://doi.org/10.1016/j.semcdb.2016.07.029>
- Mobasser SA, Zijl S, Salameti V, Walko G, Stannard A, Garcia-Manyes S, Watt FM (2019) Patterning of human epidermal stem cells on undulating elastomer substrates reflects differences in cell stiffness. *Acta Biomater* 87:256–264. <https://doi.org/10.1016/j.actbio.2019.01.063>
- Mongera A et al (2018) A fluid-to-solid jamming transition underlies vertebrate body axis elongation. *Nature* 561:401–405. <https://doi.org/10.1038/s41586-018-0479-2>
- Nagel AJLSR (1998) Jamming is not just cool any more. *Nature* 396
- Park JA, Fredberg JJ (2016) Cell jamming in the airway epithelium. *Ann Am Thorac Soc* 13(Suppl 1):S64–S67. <https://doi.org/10.1513/AnnalsATS.201507-476MG>
- Park JA et al (2015) Unjamming and cell shape in the asthmatic airway epithelium. *Nat Mater* 14:1040–+. <https://doi.org/10.1038/NMAT4357>
- Park JA, Atia L, Mitchel JA, Fredberg JJ, Butler JP (2016) Collective migration and cell jamming in asthma, cancer and development. *J Cell Sci*. <https://doi.org/10.1242/jcs.187922>
- Pegoraro AF, Fredberg JJ, Park JA (2016) Problems in biology with many scales of length: cell-cell adhesion and cell jamming in collective cellular migration. *Exp Cell Res* 343:54–59. <https://doi.org/10.1016/j.yexcr.2015.10.036>
- Petitjean L, Reffay M, Grasland-Mongrain E, Poujade M, Ladoux B, Buguin A, Silberzan P (2010) Velocity fields in a collectively migrating epithelium. *Biophys J* 98:1790–1800. <https://doi.org/10.1016/j.bpj.2010.01.030>
- Poujade M et al (2007) Collective migration of an epithelial monolayer in response to a model wound. *Proc Natl Acad Sci U S A* 104:15988–15993. <https://doi.org/10.1073/pnas.0705062104>
- Rakyan VK, Blewitt ME, Druker R, Preis JJ, Whitelaw E (2002) Metastable epialleles in mammals. *Trends Genet* 18:348–351
- Rawles ME (1947) Origin of pigment cells from the neural crest in the mouse embryo. *Physiol Zool* 20:248–266
- Rorth P (2012) Fellow travellers: emergent properties of collective cell migration. *EMBO Rep* 13:984–991. <https://doi.org/10.1038/Embor.2012.149>
- Sabass B, Gardel ML, Waterman CM, Schwarz US (2008) High resolution traction force microscopy based on experimental and computational advances. *Biophys J* 94:207–220. <https://doi.org/10.1529/biophysj.107.113670>
- Sadati M, Taheri Qazvini N, Krishnan R, Park CY, Fredberg JJ (2013) Collective migration and cell jamming. *Differentiation* 86:121–125. <https://doi.org/10.1016/j.diff.2013.02.005>
- Sadati M, Nourhani A, Fredberg JJ, Taheri Qazvini N (2014) Glass-like dynamics in the cell and in cellular collectives. *Wiley Interdiscip Rev Syst Biol Med* 6:137–149. <https://doi.org/10.1002/wsbm.1258>
- Saw TB et al (2017) Topological defects in epithelia govern cell death and extrusion. *Nature* 544:212–216. <https://doi.org/10.1038/nature21718>
- Scarpa E, Mayor R (2016) Collective cell migration in development. *J Cell Biol* 212:143–155. <https://doi.org/10.1083/jcb.201508047>
- Schwarz US, Soine JR (2015) Traction force microscopy on soft elastic substrates: A guide to recent computational advances. *Biochim Biophys Acta* 1853:3095–3104. <https://doi.org/10.1016/j.bbamcr.2015.05.028>
- Sepulveda N, Petitjean L, Cochet O, Grasland-Mongrain E, Silberzan P, Hakim V (2013) Collective cell motion in an epithelial sheet can be quantitatively described by a stochastic interacting particle model. *PLoS Comput Biol* 9:e1002944. <https://doi.org/10.1371/journal.pcbi.1002944>
- Sixt M, Engelhardt B, Pausch F, Hallmann R, Wendler O, Sorokin LM (2001) Endothelial cell laminin isoforms, laminins 8 and 10, play decisive roles in T cell recruitment across the blood-brain barrier in experimental autoimmune encephalomyelitis. *J Cell Biol* 153:933–946. <https://doi.org/10.1083/jcb.153.5.933>
- Song J et al (2017) Endothelial basement membrane laminin 511 contributes to endothelial junctional tightness and thereby inhibits leukocyte transmigration. *Cell Rep* 18:1256–1269. <https://doi.org/10.1016/j.celrep.2016.12.092>
- Steinberg MS (2007) Differential adhesion in morphogenesis: a modern view. *Curr Opin Genet Dev* 17:281–286. <https://doi.org/10.1016/j.gde.2007.05.002>
- Sunyer R et al (2016) Collective cell durotaxis emerges from long-range intercellular force transmission. *Science* 353:1157–1161. <https://doi.org/10.1126/science.aaf7119>
- Sutherland HG, Kearns M, Morgan HD, Headley AP, Morris C, Martin DI, Whitelaw E (2000) Reactivation of heritably silenced gene expression in mice. *Mamm Genome* 11:347–355
- Tambe DT et al (2011) Collective cell guidance by cooperative intercellular forces. *Nat Mater* 10:469–475. <https://doi.org/10.1038/NMAT3025>
- Tarle V, Ravasio A, Hakim V, Gov NS (2015) Modeling the finger instability in an expanding cell monolayer. *Integr Biol (Camb)* 7:1218–1227. <https://doi.org/10.1039/c5ib00092k>
- Trappe V, Prasad V, Cipelletti L, Segre PN, Weitz DA (2001) Jamming phase diagram for attractive particles. *Nature* 411:772–775. <https://doi.org/10.1038/35081021>
- Trepat X, Fredberg JJ (2011) Plithotaxis and emergent dynamics in collective cellular migration. *Trends Cell Biol* 21:638–646. <https://doi.org/10.1016/j.tcb.2011.06.006>
- Trepat X, Sahai E (2018) Mesoscale physical principles of collective cell organization. *Nat Phys* 14:671–682. <https://doi.org/10.1038/s41567-018-0194-9>
- Trepat X, Wasserman MR, Angelini TE, Millet E, Weitz DA, Butler JP, Fredberg JJ (2009) Physical forces during collective cell migration. *Nat Phys* 5:426–430. <https://doi.org/10.1038/nphys1269>
- Vishwakarma M, Di Russo J, Probst D, Schwarz US, Das T, Spatz JP (2018) Mechanical interactions among followers determine the emergence of leaders in migrating epithelial cell collectives. *Nat Commun* 9:3469. <https://doi.org/10.1038/s41467-018-05927-6>
- Wagstaff L, Kolahgar G, Piddini E (2013) Competitive cell interactions in cancer: a cellular tug of war. *Trends Cell Biol* 23:160–167. <https://doi.org/10.1016/j.tcb.2012.11.002>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.